



Pesticide Resistance Strategy

New Zealand Plant Protection Society



- [Home](#)
 - [Introduction](#)
 - [Site map](#)
 - [New Zealand Committee on Pesticide Resistance \(NZCPR\)](#)
 - [New Zealand Plant Protection Society website](#)
- [Herbicides](#)
 - [Introduction](#)
 - [Modes of action](#)
 - [Resistance management principles](#)
 - [Triazine in maize](#)
 - [Triazine in arable crops](#)
 - [Triazine in orchards](#)
 - [Phenoxy](#)
 - [Picolinic](#)
 - [Sulfonylurea](#)
 - [Glyphosate](#)
- [Insecticides & miticides](#)
 - [Introduction](#)
 - [Modes of action](#)
 - [Resistance management principles](#)
 - [Blowflies](#)
 - [Diamondback moth](#)
 - [Green peach aphid](#)
 - [Leafhoppers](#)
 - [Leafrollers](#)
 - [Lettuce aphid](#)
 - [Mealybugs](#)
 - [Melon aphid](#)
 - [Sheep biting-louse](#)
 - [Spider mite](#)
 - [Thrips](#)

- [Tomato fruitworm](#)
- [Whitefly](#)
- [Fungicides & bactericides](#)
 - [Introduction](#)
 - [Modes of action](#)
 - [Resistance management principles](#)
 - [Anilinopyrimidines](#)
 - [Anilinopyrimidines in grapes](#)
 - [Benzimidazoles](#)
 - [Carboxylic acid amides](#)
 - [Demethylation inhibitors](#)
 - [Dicarboximides](#)
 - [Dodine](#)
 - [Morpholines](#)
 - [Phenylamides](#)
 - [Qo inhibitors](#)
 - [Succinate dehydrogenase inhibitors](#)
 - [Streptomycin](#)
- [Crops](#)
 - [Introduction](#)
 - [Apples - black spot](#)
 - [Grapes - botrytis bunch rot](#)
 - [Grapes - powdery mildew](#)
 - [Maize - weeds](#)
 - [Summerfruit - diseases](#)

Sheep biting-louse resistance management



Sheep biting-louse (*Bovicola ovis*) has developed resistance to synthetic pyrethroid insecticides
(Photo: Michel Valim, www.phthiraptera.pop.com.br)

A.C.G. Heath
AgResearch Wallaceville, PO Box 40063, Upper Hutt, New Zealand

(Revised October 2004)

Reason for strategy

The sheep biting-louse, *Bovicola ovis* (formerly *Damalinia ovis*), has developed resistance to the synthetic pyrethroid cypermethrin, and side resistance to other members of that chemical family (Levot et al. 1995; Wilson et al. 1997). Management strategies aimed at reducing reliance on insecticides will not only conserve the efficacy of existing products, but will help reduce residues in wool to levels that are acceptable to overseas markets.

Background

The sheep biting-louse, *B. ovis*, has been present in New Zealand since the first days of sheep farming and, as a consequence, the louse has been exposed to a wide range of pesticide families. Fortunately there appears to have been no ability on the part of the louse to develop resistance until synthetic pyrethroids were introduced in the late 1970s. Even then there was no sign of reduced efficacy until about the early 1990s (Levot 1992; Wilson et al. 1997). A reduced susceptibility to organophosphates has also been detected in Australia (Levot 1994).

The biting-louse is widespread in New Zealand sheep flocks and can downgrade wool quality (Kettle & Pearce 1974), but more importantly, can reduce lamb pelt quality significantly because of cockle, the manifestation of the sheep's immune response to lice (Heath et al. 1994; Pfeffer et

al. 1997). In recent years, insecticide residues in wool have posed a potential trade barrier as European markets try to protect the environment and water quality downstream from wool-scouring plants (Edwards 1998).

Because sheep dipping is not always a straightforward or cheap procedure, farmers often try and facilitate it and improve cost-effectiveness, by using insecticides that combine flystrike prevention with louse control in one operation. This means that selection pressure for resistance can operate on both parasites simultaneously (Sales et al. 1996). This is not a desirable outcome. Unfortunately, only a small number of products with single species label claims exist, so choice is limited. Simultaneous selection can be avoided if dipping to control lice is done outside the blowfly activity period, because lice do not leave the host and are available to insecticides all year round.

Products with label claims for control of sheep biting-lice in New Zealand

Table 1: Products with label claims for control of sheep biting-lice in New Zealand (Nottingham 2003). Many of these products have a dual purpose and are used for preventing flystrike as well, whereas others have even wider claims that include many sheep ectoparasites. These label claims are not shown here. Method of application varies with products and is shown for each as the choice of high versus low volume can be relevant in resistance management and environmental protection.

Pesticide category and IRAC chemical group	Method of application
Pesticide common and product names	
Organophosphate 1B	
chlorfenvinphos (Supreme)	Plunge, shower, jetting
chlorpyrifos (Xterminate 10)	Plunge, shower, jetting
coumaphos (Asuntol liquid)	Plunge, shower
propetamphos (eNkamphos 500, eNkamphos 1250)	Plunge, shower, jetting
Organophosphate/pyrethroid 1B/3	
chlorpyrifos/cypermethrin (Flypel)	Applicator/spray
Pyrethroid 3	
Alpha-cypermethrin (Duracide, Vanquish)	Pour-on
cypermethrin (Cypercure) ¹	Applicator/pour-on
High- <i>cis</i> cypermethrin (Avalanche)*	Applicator/spray
High- <i>cis</i> cypermethrin (Cypor)	Applicator/pour-on
High- <i>cis</i> cypermethrin (Ectomin 100 EC)*	Plunge, shower
deltamethrin (Wipe-out) ¹	Applicator/pour-on
Cyano-pyrethroid 3	
Cyhalothrin (Grenade) ¹	Plunge, shower
Macrocyclic lactone (avermectin/milbemycin) 6	

ivermectin (Jetamec jetting fluid for sheep)	Jetting
ivermectin (Erase MPC)	Spray/jetting
Benzoyl urea 15	
diflubenzuron (Blitz, Ectogard, Fleecemaster, Zenith)	Plunge, shower, jetting
diflubenzuron (Magnum, Zenith spray on)	Applicator/spray
triflumuron (Zapp, Zapp jetting liquid)	Applicator/spray, jetting
Formamidine 19	
amitraz (Taktic) ¹	
Spinosyn 5	
spinosad (Extinosad)	Plunge, shower, jetting

¹For flystrike prevention only; no biting-louse claim.

Current status of louse resistance in New Zealand

Synthetic pyrethroid-resistant louse populations occur throughout New Zealand, especially those resistant to cypermethrin-based formulations, with a strong likelihood of side resistance to other pyrethroids (Wilson et al. 1997). There have been no reports from the field of reduced efficacy with organophosphates, although some evidence from Australia suggests that such may be occurring (Levot 1994). All other chemical groups remain efficacious.

Resistance management and prevention strategy

Because all chemical classes, apart from pyrethroids, are fully efficacious, louse control is not difficult to achieve. Nevertheless, in an attempt to reduce the risk of selection pressure, rotation of chemical classes may be advantageous. An important principle, however, is to direct farmers towards dipping to control lice only if the sheep are infested. Many farmers dip on an annual basis as a form of insurance, whether or not their sheep are infested. A simple inspection procedure can determine whether sheep require dipping, but there is also a technology based on the ELISA test which can detect lice based on the presence of louse-specific proteins deposited in wool in louse faeces (Sanderson 2001; A.T. Pfeffer, pers. comm.).

Another good management procedure is to treat sheep immediately off-shears, which ensures a greater proportion of lice come into contact with insecticide, and reduces the volume of dip chemical necessary to achieve this. Wool growth then ensures that the half-life of chemicals is reduced, with consequent reduced selection pressure.

If ewes are dipped during early pregnancy, the risk that they still will be louse-infested at lambing is reduced, as is the chance of lambs becoming infested. Dipping the young lambs results in the flock getting as close to louse-free as possible. The ewe lambs, when old enough to be mated, should not then be in a position to so readily infest their progeny. This could lead to a situation where dipping is only necessary every alternate year, or even at longer intervals,

assuming that there is a residual louse population, something that is quite common with the vagaries inherent in dipping.

A good degree of louse control can be achieved by shearing only because solar radiation and dehydration reduce the hatchability of louse eggs. These effects can be enhanced by dipping newly shorn sheep in a water/detergent mix, or with naturally occurring insecticides (Heath et al. 1995), or with conventional products which will provide a better measure of louse control than if they were applied to long-wooled sheep.

Note: Control failure does not always imply resistance

The following guidelines are recommended:

- Use dips as soon as possible after shearing to achieve maximum efficacy.
- Try and dip sheep during months when blowflies are not active.
- Only dip sheep if they are louse-infested; use direct inspection or a louse-detection kit.
- Ensure that manufacturers' recommendations are scrupulously followed and that all apparatus is calibrated correctly and working effectively
- Follow good husbandry and integrated management procedures as described in Edwards et al. (2001).

References

Edwards S 1998. Wools of New Zealand approach to dip residue management. Proceedings of the 28th Seminar of the Society of Sheep and Cattle Veterinarians, NZVA. Publication No. 180. Veterinary Continuing Education, Massey University, Palmerston North. Pp. 1-7.

Edwards S, Marshall A, Cole D, Heath ACG 2001. Fly and Lice, Numbering their Days. WoolPro, Wellington, New Zealand. 52 p.

Heath ACG, Cole DJW, Bishop DM, Pfeffer A, Cooper SM, Risdon P 1994. Preliminary investigations into the aetiology and treatment of cockle, a sheep pelt defect. *Veterinary Parasitology* 56: 239-254.

Heath ACG, Lampkin N, Jowett JH 1995. Evaluation of non-conventional treatments for control of the biting louse (*Bovicola ovis*) on sheep. *Medical and Veterinary Entomology* 9: 407-412.

Kettle PR, Pearce DM 1974. Effect of the sheep body louse (*Damalinia ovis*) on host weight gain and fleece value. *New Zealand Journal of Experimental Agriculture* 2: 219-221.

Levot GW 1992. High-level resistance to cypermethrin in the sheep body louse, *Damalinia ovis* (Schränk). *Australian Veterinary Journal* 69: 120.

Levot GW 1994. A survey of organophosphate susceptibility in populations of *Bovicola ovis* (Schränk) (Phthiraptera: Trichodectidae). *Journal of the Australian Entomological Society* 33: 31-34.

Levot GW, Johnson PW, Hughes PB, Powis KJ, Boray JC, Dawson KL 1995. Pyrethroid resistance in Australian field populations of the sheep body louse, *Bovicola (Damalinia) ovis*. *Medical and Veterinary Entomology* 9: 59-65.

Nottingham R ed. 2003. IVS Manual 2003, Volume 18. MediMedia, Auckland, New Zealand.

IVS Annual 2005. www.ivsonline.co.nz

Pfeffer AT, Phegan MD, Bany J 1997. Detection of homocytotropic antibody in lambs infested with the louse, *Bovicola ovis*, using a basophil histamine-release assay. *Veterinary Immunology and Immunopathology* 57: 315-325.

Sales N, Shivas M, Levot G 1996. Toxicological and oviposition suppression responses of field populations of the Australian sheep blowfly, *Lucilia cuprina* (Wiedmann) (Diptera:Calliphoridae) to the pyrethroid cypermethrin. *Australian Journal of Entomology* 35: 285-288.

Sanderson L 2001. Checking for lice like a pregnancy test. *Wool Grower* No. 11, spring 2001:13.

Wilson JA, Heath ACG, Quilter S, McKay C, Litchfield D, Nottingham R 1997. A preliminary investigation into resistance to synthetic pyrethroids by the sheep biting louse (*Bovicola ovis*) in New Zealand. *New Zealand Veterinary Journal* 45: 8-10.

[Site map](#) · [Author's login](#)



© 2020 New Zealand Plant Protection Society (Incorporated). All rights reserved.

The New Zealand Plant Protection Society (Incorporated) shall not be liable for the commercial performance of any products or any losses arising from the use of the information contained herein.