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Description of the postembryonic stages of *Mulcticola macrocephalus* (Kellogg, 1896) (Phthiraptera, Ischnocera: Philopteridae)

Kamila M.D. Kuabara¹ , Michel P. Valim² , and Terry D. Galloway^{3*} 

¹The Academy of Natural Sciences of Drexel University, Philadelphia, Pennsylvania, United States of America, ²Universidade Iguaçu, Nova Iguaçu, Rio de Janeiro, Brazil and ³Department of Entomology, University of Manitoba, Winnipeg, Manitoba, R3T 2N2, Canada

*Corresponding author. Email: terry.galloway@umanitoba.ca

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Abstract

The nymphs I, II, III, and adult male and adult female stages of *Mulcticola macrocephalus* (Kellogg) (Phthiraptera, Ischnocera: Philopteridae), a monoxenous species of chewing louse infesting common nighthawk, *Chordeiles minor* (Forster) (Caprimulgiformes: Caprimulgidae), in North America, are described. Line drawings of whole nymphs and adults are presented, as are details of anterior dorsal head plates, metasternal plates, female subvulvar plates, and male genitalia. Ontogenetic characters exhibited during the postembryonic development of this species of chewing louse are described and discussed in relation to previous information in the literature.

Introduction

Lice in the genus *Mulcticola* infest members of the Caprimulgidae (Aves: Caprimulgiformes), the nighthawks and nightjars (Price *et al.* 2003). This group of chewing lice includes 18 species, infesting 24 host species, in 25 countries (Valim and Kuabara 2015). The genus has attracted recent attention taxonomically (Valim and Kuabara 2015) and ecologically (Galloway 2006; Galloway and Lamb 2015). Valim and Kuabara (2015) reviewed the neotropical species, described five new species, and provided a catalogue for all *Mulcticola* species, while Galloway (2006) and Galloway and Lamb (2015) calculated infestation parameters (prevalence and intensity) and population variability for one species, *Mulcticola macrocephalus* (Kellogg) (Phthiraptera, Ischnocera: Philopteridae), infesting the common nighthawk, *Chordeiles minor* (Forster) in Manitoba, Canada. In the course of the ecological studies in Manitoba, large numbers of lice were collected, including all three immature stages. This species of chewing louse was originally described by Kellogg (1896), but many descriptive details for *M. macrocephalus* were not provided at that time. The abundance of specimens collected from common nighthawks in Manitoba offered the opportunity to redescribe the adults of *M. macrocephalus*, as well as the three juvenile instars.

Material and methods

Lice were collected from common nighthawks that had been submitted to two wildlife rehabilitation hospitals in Manitoba, the Wildlife Haven (previously Manitoba Wildlife Rehabilitation Organization) and the Prairie Wildlife Rehabilitation Centre. Nighthawks that died or were

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euthanised were individually frozen at -20°C for at least 48 hours to kill all ectoparasites. Birds were washed three times to remove ectoparasites, and samples were passed through a $90\text{-}\mu$ sieve. Lice were handpicked from samples and preserved in 70% or 95% ethanol. All specimens from the current descriptive study are deposited in the Museu de Zoologia da Universidade de São Paulo (MZUSP), Brazil, with a number associated as a unique identifier from the SophiA® database for the Phthiraptera collection. Additional specimens are deposited in the J.B. Wallis/R.E. Roughley Museum of Entomology, Department of Entomology, University of Manitoba, Winnipeg, Manitoba, Canada. Lice were permanently slide mounted in Canada balsam, following the technique in Palma (1978). The terminology of body features and chaetotaxy follows that used by Valim and Kuabara (2015; see also Fig. 4).

All measurements are in millimetres and identified by the following abbreviations: ADHPL – anterior dorsal head plate length; ADHPW – anterior dorsal head plate width; AW – abdominal width (at the level of segment V); GL – male genitalia length; HL – head length (at midline and including the hyaline margin); PAW – preantennal width (at the level of preconal setae); PL – paramere length; PTW – pterothorax width (at its posterior level); PW – prothorax width; TL – total length; TW – temporal width (at the level of marginal temporal setae 2 – *mts2*). The cephalic index measure is calculated by the head length divided by head width.

Systematics

Phthiraptera Haeckel, 1896

Ischnocera Kellogg, 1896

Philopteridae Burmeister, 1838 (*sensu lato*)

Mulcticola Clay et Meinertzhagen, 1938, p. 279

Mulcticola macrocephalus (Kellogg, 1896) (See Figs. 1–4)

Lipeurus macrocephalus Kellogg 1896, p. 504, type host *Chordeiles virginianus henryi* (= *C. minor henryi*).

Nirmus tyrannus: Osborn 1896, p. 228, ex *Tyrannus* sp. (= error) (junior synonym, see Emerson 1960, p. 162).

Mulcticola macrocephala: Carriker 1947, p. 136 (new combination and unjustified emendation).

Mulcticola macrocephalus: Hopkins and Clay 1952, p. 227 (checklist); Carriker 1957, p. 103 (lectotype designation); Emerson and Elbel 1957, p. 423 (illustrations of anterior dorsal head plate and male genitalia); Emerson 1972, p. 96 (list of North American species); Price *et al.* 2003, p. 198 (checklist); Galloway 2006, p. 7 (biological and parasitological indices); Valim and Kuabara 2015, p. 12 (catalogue of species); Galloway and Lamb 2015, p. 725 (ecology).

Type locality: Palo Alto, California, United States of America

Descriptions of preimaginal stages

Nymph I. General habitus as in Fig. 1A; details of head and thoracic chaetotaxy as in Fig. 2A; cephalic ratio 1.3–1.4, with marginal carina medially interrupted, region smooth and distinctly sclerotised, without an anterior dorsal head plate. Without visible attachment sites for cibarial muscles on the head capsule. The *as3* present, hyaline margin completely absent; *pas*, *os*, *mts1* and *mts3* very long; *ads* set on sclerotised part anterior to ecdysial line. Prothorax with postspiracular setae very long, reaching posterior margin of pterothorax; chaetotaxy of pterothorax, each side: one short, spine-like posterolateral seta plus one long posterior seta. Meso- and metasternal plates without sclerotisation or setae. Abdomen membranous, without sclerotised plates. Tergal chaetotaxy (each side): II with one anterior seta; II–VII, IX with one posteromedial seta; VIII with one short and one long seta; IV–VIII with postspiracular setae. Sternal chaetotaxy (each side): II–VIII

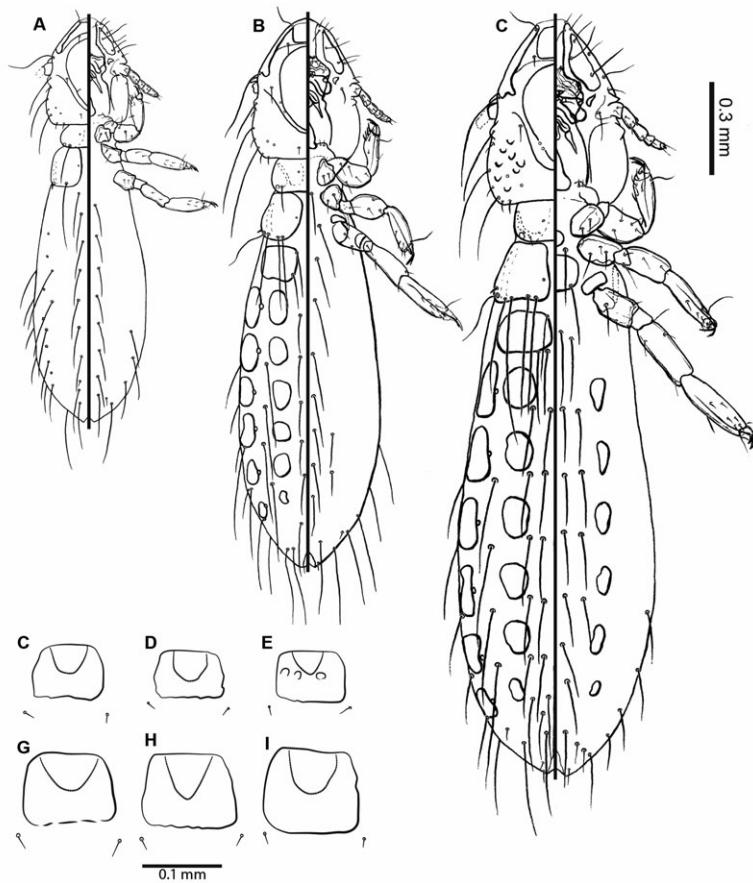


Fig. 1. Nymphal habitus of *Multicola macrocephalus*, in dorsoventral views: **A**, nymph I; **B**, nymph II; **C–E**, anterior dorsal head plate from three specimens of nymph II; **F**, nymph III; **G–I**, anterior dorsal head plate from three specimens of nymph III.

with one medial seta; IX with one long and one short seta. Pleural chaetotaxy (each side): IV–V one seta; VI–VII two setae; IX–X one seta. Measurements ($n = 9$): HL 0.33–0.35; PAW 0.21–0.24; TW 0.25–0.27; PW 0.16–0.20; PTW 0.19–0.25; AW 0.23–0.33; TL 1.00–1.29.

Nymph II. General habitus as in Fig. 1B; details of head and thoracic chaetotaxy as in Fig. 2B; cephalic ratio 1.3, with marginal carina medially interrupted and developed anterior dorsal head plate posteriorly still fused with the head surface. Three variations of the anterior dorsal head plate are presented in Fig. 1C–E. Attachment sites for cibarial muscles indistinct, aspect of anterior dorsal head plate smooth. The *as3* present and set on hyaline margin; *pas*, *os*, *mts1* and *mts3* very long; *ads* set on sclerotised part of head posterior to the anterior dorsal head plate. Prothorax (each side) with postspiracular setae intermediate between very long in nymph I and very short in nymph III; chaetotaxy of pterothorax, each side: one spine-like ventral seta, one trichoid lateral seta, one long posterolateral seta, plus two long posteromedial setae. Chaetotaxy of pterothorax similar to that of imago, less one long seta on posterolateral margin of metathorax. Meso- and metasternal plates without sclerotisation but with one pair of medium-long setae on each segment. Abdomen membranous but with sclerotised pleural and tergal plates. Segment II with one subquadrate tergopleural plate; segments III–VIII with distinct, square-shaped tergal plates fused with pleural ones. Tergal chaetotaxy (each side): II with one anterior seta; II–VIII with one postero-medial seta; IV–VII with postspiracular setae; IX with two long setae. Sternal chaetotaxy (each

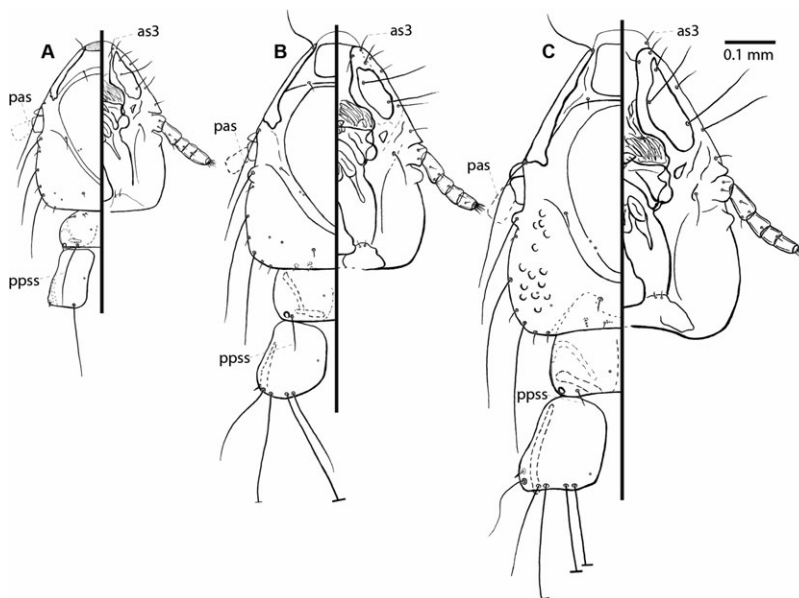


Fig. 2. Head in dorsoventral views and thorax in dorsal view of *Multicola macrocephalus*: **A**, nymph I; **B**, nymph II; **C**, nymph III. *as*, anterior seta; *pas*, preantennal seta; *ppss*, prothoracic postspiracular seta.

side): II–VIII with one medial seta; VI–VII with additional pair of setae; IX with two long setae. Pleural chaetotaxy (each side): IV–V one seta; VI–VIII two setae; IX–X one seta. Measurements ($n = 6$): HL 0.39–0.46; PAW 0.27–0.32; TW 0.30–0.36; ADHPW 0.08–0.09; PW 0.20–0.26; PTW 0.29–0.36; AW 0.34–0.46; TL 1.46–1.79.

Nymph III. General habitus as in Fig. 1F; details of head and thoracic chaetotaxy as in Fig. 2C; cephalic index 1.3, with marginal carina medially interrupted and developed anterior dorsal head plate, posteriorly fused with the head surface. Three variations of the anterior dorsal head plate are presented in Fig. 1G–I. Attachment sites of cibarial muscles clearly discernible on the anterior dorsal head plate. The *as3* present and set on hyaline margin; *pas*, *os*, *mts1* and *mts3* very long; *ads* set on the soft integument of preantennal suture, posterior to the anterior dorsal head plate. Prothorax with postspiracular setae very short, as in imago; chaetotaxy of pterothorax as in imago, with four long posterior setae plus spine-like and trichoid setae. Meso- and metasternal plates with sclerotisation and with one pair of medium-long setae on each segment. Abdomen membranous but with sclerotised pleural, tergal, and sternal plates. Tergal plates as described for nymph II, sternal plates tear-shaped and present on segments III–VIII. Tergal chaetotaxy (each side): II with one anterior seta; II–IX with two posteromedial setae; VIII with one additional seta; IV–VII with postspiracular setae. Sternal chaetotaxy (each side): II–VI with two medial setae; VII–VIII with one seta each; X with one long, two short, plus one long setae. Pleural chaetotaxy (each side): IV–V one seta; VI–VII two setae; VIII three (rarely two) setae; IX–X two setae. Measurements ($n = 7$): HL 0.54–0.58; PAW 0.37–0.42; TW 0.42–0.46; ADHPW 0.10–0.11; PW 0.29–0.32; PTW 0.39–0.43; AW 0.44–0.61; TL 1.94–2.37.

Redescription of the imago

Head. Triangular, sub-truncate anteriorly, longer than wide, cephalic index 1.2 in both sexes, rarely 1.3 (Fig. 4A). The short clypeal region, narrow hyaline margin, rounded sides, and slightly rounded to straight in front. Anterior dorsal and ventral head plates of characteristic shape, with

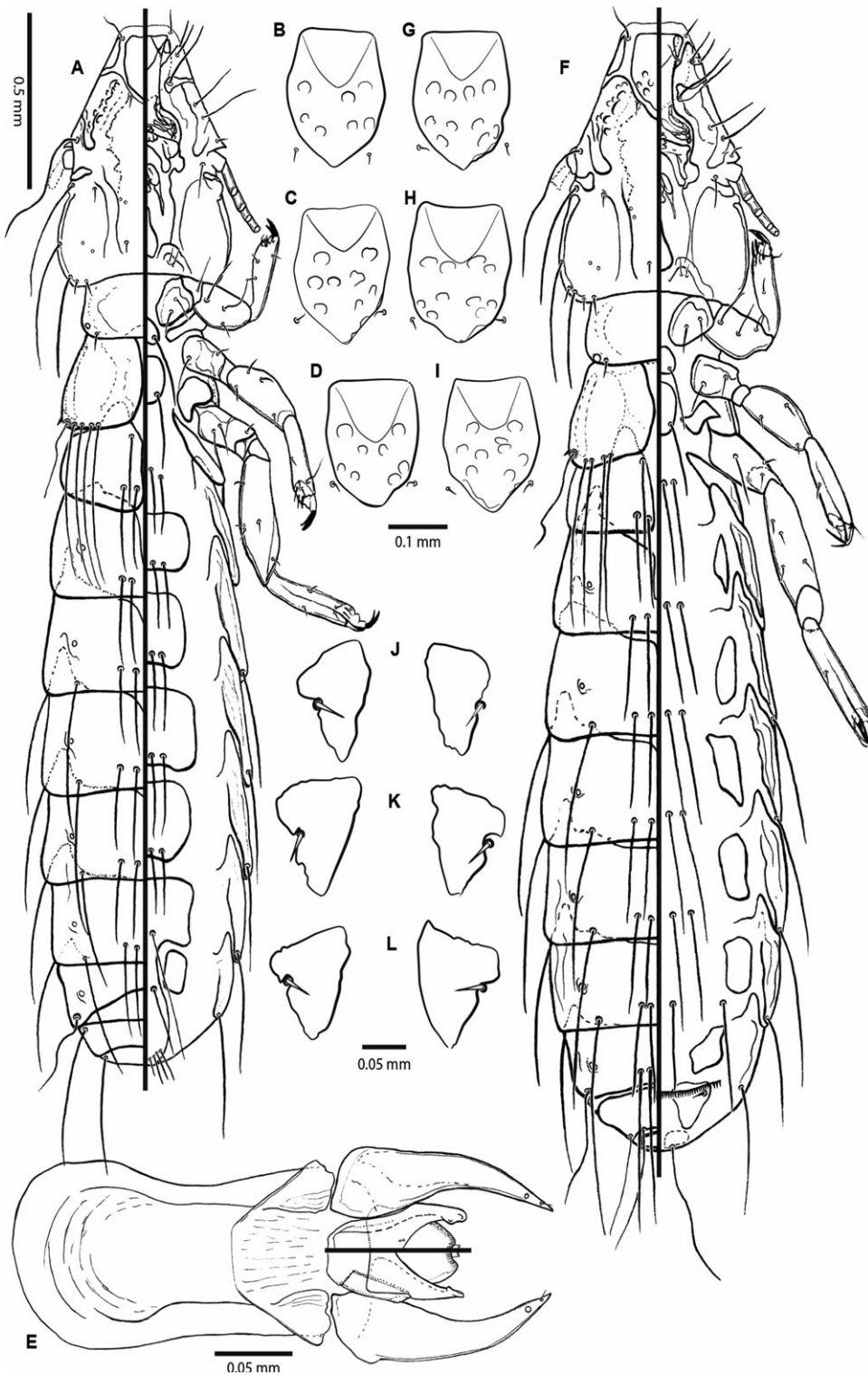


Fig. 3. Adults of *Multicola macrocephalus* (Kellogg, 1896) in dorsoventral views: **A**, male habitus; **B–D**, anterior dorsal and ventral head plates of males from three specimens; **E**, male genitalia with mesosome; **F**, female habitus; **G–I**, anterior dorsal and ventral head plates of females; **J–L**, female subvulvar plates from three specimens.

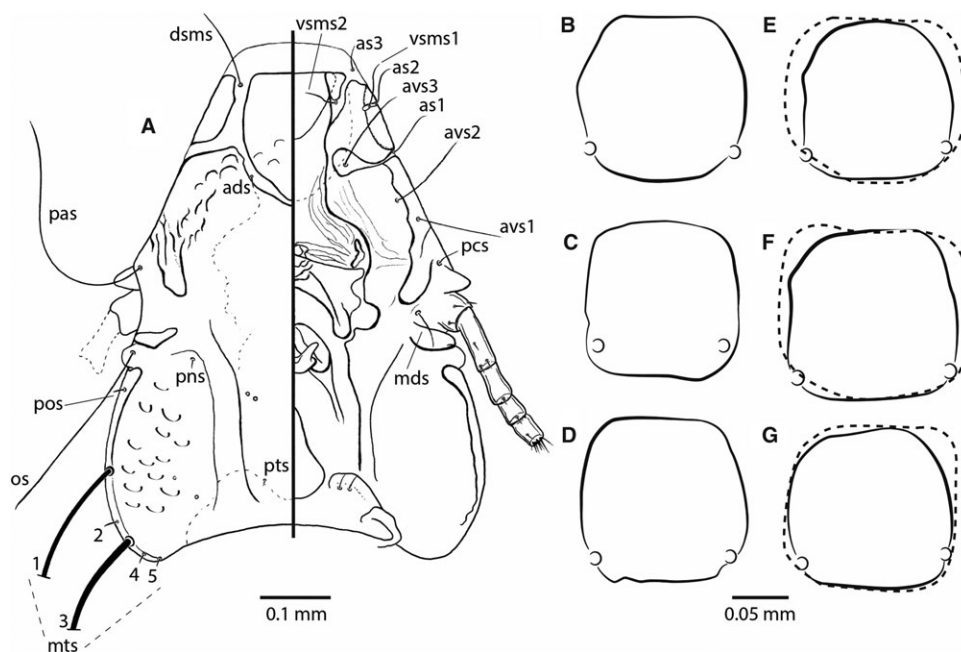


Fig. 4. *Mulcticola macrocephalus* (Kellogg, 1896): **A**, male head in dorsoventral views; **B–D**, female metasternal plates from three specimens; variation of three specimens; **E–G**, male metasternal plates from three specimens, *Mulcticola nacunda* Carriker, 1945 (dotted lines). *ads*, anterior dorsal seta; *as*, anterior seta; *avs*, anterior ventral seta; *dsms*, dorsal submarginal seta; *mds*, mandibular seta; *mts*, marginal temporal seta; *os*, ocular seta; *pas*, preantennal seta; *pcs*, preconal seta; *pns*, postnodal seta; *pos*, preocular seta; *pts*, posttemporal seta; *vsm*, ventral submarginal seta.

distinct attachment sites for cibarial muscles on dorsal plate (Fig. 4A); the ratio of anterior head plate 1.3, rarely 1.2 in some females. Coni triangular in shape, short, not reaching the posterior margin of scapus. Postnodal seta short. Eye small, rounded, and prominent. Temples slightly extended, marginal temporal carina narrow. Posterior margin of occipital edge concave, with *pts* and sensilla *s1–s4* present on each side.

Thorax. Entire thorax is shorter than the head. Prothorax trapezoidal, pterothorax also as a larger trapezium, about the same width as the head. Meso- and metasternal plates distinctly sclerotised, each with one pair of medium-long setae, latter plate of diagnostic shape, nearly square (Fig. 4B–G).

Abdomen. Elongated, with the widest segment at V (Fig. 3A, F). The tergal plate of segment II laterally fused with pleura and entirely divided medially, with four posterior setae, plus one pair of anterior setae. Segments III–VII medially entire and fused to pleura laterally, forming a tergo-pleurite. Postspiracular setae set dorsally each side on IV–VII, segment VIII with trichoid setae set posterolaterally; sensilla accessory to postspiracular setae on IV–V. Posterior setae on tergo-pleurites III–VII 4 (rarely 5) and VIII 4 (rarely 2–3); anterior margin of tergal plates III–V (6) or III–VI (6) with a median notch, gradually narrowing into posterior segments. Tergite VIII fused medially, but in males medially narrower than in III–VII. Pleural bands extended into the preceding segment, forming well-defined “re-entrant heads” on III–VIII; pleural chaetotaxy: 0 on II–III; 2 (rarely 1 in one side) setae on IV–V; 3 (rarely 2) setae on VI–VIII. Sternal chaetotaxy: II with 4 (rarely 3 or 5) setae; III–VI with 3–6 setae (but mode and median 4 setae). Sternites dimorphic, ovoid, and set laterally on each side of abdominal segments III–VII in females and roughly rectangular in shape and set across each segment III–VI in males.

Male. General habitus as in Fig. 3A. Subgenital plate with 4 (rarely 5) setae. Copulatory apparatus of male (Fig. 3E) typical for genus; endomeral plate broad, its lateral arms characteristic (Fig. 3E) with pointed tips dorsally and slightly broad on ventral side. Ventrally one pair of small sensilla medial on each side of sclerotised part of endomera. Measurements ($n = 3$): HL 0.68–0.72; PAW 0.45–0.48; TW 0.55–0.60; ADHPL 0.19–0.21; ADHPW 0.16–0.17; PW 0.38–0.41; PTW 0.50–0.53; AW 0.64–0.74; GL 0.34; PL 0.13–0.14; TL 2.93–3.08.

Female. General habitus as in Fig. 3F. One pair of female subvulvar sclerites well developed and of characteristic shape, with one spine-like marginal seta on each plate (Fig. 3J–L). Subgenital plate with 4–6 setae each side (but mode and median 6 setae); vulvar margin with 12–19 thin and 14–27 spine-like setae, plus 10–17 small setae anterior to the vulvar margin. Measurements ($n = 9$): HL 0.67–0.76; PAW 0.44–0.50; TW 0.55–0.63; ADHPL 0.18–0.22; ADHPW 0.14–0.18; PW 0.39–0.44; PTW 0.51–0.58; AW 0.66–0.77; TL 2.87–3.23.

Material examined: Canada. Manitoba: 2♂, 6 NI, 3 NII, 5 NIII (MZUSP 7585–7596), ex *Chordeiles minor minor* (Forster, JR, 1771) (case no.: CONI/1257/CEN/02), Winnipeg (49°53'24" N, 97°8'36.96" W), 6.IX.2002, coll. T.D. Galloway/D. Holder. 5♀, 3 NI, 2 NII, 1 NIII (MZUSP 7597–7611), ex *C. m. minor* (CONI/1280/CEN/02), Winnipeg (49°53'24" N, 97°8'36.96" W), 10.IX.2002, coll. T.D. Galloway/D. Holder. 1♂, 4♀, 1 NII, 1 NIII (MZUSP 7612–7618), ex *C. m. minor* (CONI/819/CEN/14), Grande Pointe (49°46'0.12" N, 97°3'2.30" W), 21.VIII.2014, coll. T.D. Galloway/E.N. McNally.

Remarks

Mulcticola macrocephalus is morphologically close to *M. nacunda* Carriker, 1945 by the shape of the anterior dorsal head plate, based on its dimorphic sternal plates and the chaetotaxy of abdominal tergites and sternites. They can be distinguished by several characteristics: the length-to-width ratio of the head is slightly greater in *M. macrocephalus* (1.2–1.3 versus 1.1); the metasternal plate tends to be narrower in *M. macrocephalus* (Fig. 4E–G); the inner margins of each subvulvar plate are roughly straight to slightly undulate and the antero-medial angles are blunt in females of *M. macrocephalus*, whereas in *M. nacunda*, these margins are concave and terminate in a prolonged antero-medial extension (Valim and Kuabara 2015); there are no differences in the tips of the endomeral arms in the male genitalia, at least dorsally, but the shape of the mesosome is distinctive.

Both species are similar in size, but males of *M. macrocephalus* tend to have the head (HL: 0.68–0.72 versus 0.64–0.67) and anterior dorsal plate (ADHPL: 0.19–0.21 versus 0.17–0.18) longer than in *M. nacunda*, whereas its male genitalia are smaller (GL: 0.34 versus 0.35–0.36; PL: 0.13–0.14 versus 0.16–0.17).

The third instar of *M. nacunda* was described by Valim and Kuabara (2015), so a direct comparison between the preimaginal forms of these two species is possible. Both species present the most conservative characters of the genus (see Valim and Kuabara 2015), in that they differ by the thickness of the dorsal marginal carina, being thinner in *M. macrocephalus*; the shape of the dorsal anterior head plate differs, its posterior margin being more defined and straight in *M. macrocephalus*, whereas in *M. nacunda*, there is a less sclerotised area forming a posterior medial notch. Regarding the generic features of preimagos, it is important to stress that only the nymphs II of *M. piacentinni* Valim et Kuabara and *M. bacurau* Valim et Kuabara are known. The postspiracular prothoracic setae are short in nymph III of *M. bacurau*, medium length in specimens of *M. macrocephalus* (Fig. 2B), and neither very long as in nymph I (*M. piacentinni*, *M. bacurau*, *M. nacunda*, *M. macrocephalus*) nor very short as in nymph III (aforementioned species, except *M. piacentinni*, which is unknown).

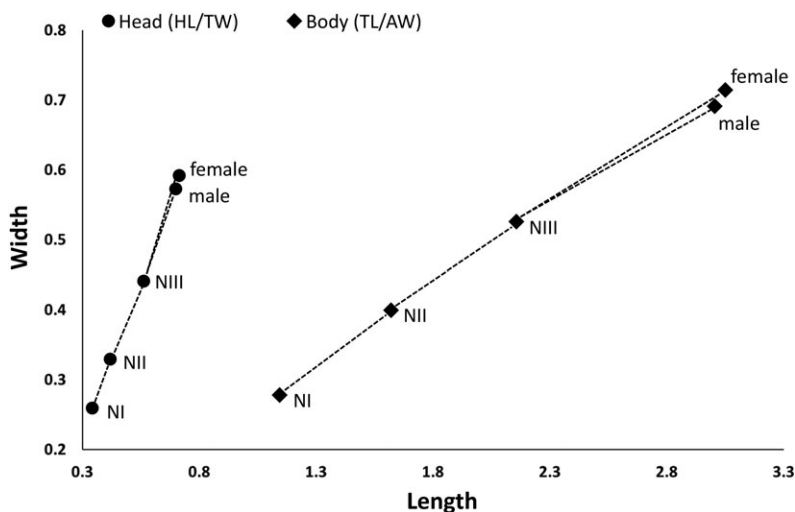


Fig. 5. Ratios between head and total lengths compared to greatest head and abdominal widths, respectively, in the postembryonic stages of *Mulcticola macrocephalus*. Males in blue; females in red after third-instar nymph. All measurements in millimetres. AW, abdominal width; HL, head length; TL, total length; TW, temporal width.

Discussion

The nymphs of lice have seldom formed the focus for descriptive analysis or subsequent application for the study of phylogenetic relationships, as has been done for some other groups of insects (e.g., the Dytiscidae (Coleoptera): Nilsson 1989; Alarie 1991). Some of the early work of Ferris (1951), Pratt and Karp (1953), and Cook and Beer (1959) led to descriptions of the nymphs of many species of sucking lice (Phthiraptera, Anoplura) by Kim (e.g., Kim 1965, 1966a, 1966b). Price and Hellenthal (1996) and Page *et al.* (1995) described the complex relationships among chewing lice (Phthiraptera: Trichodectidae) infesting pocket gophers (Rodentia: Geomyidae) in the New World using characters of first-instar nymphs of *Geomydoecus* Ewing and *Thomomydoecus* Price et Emerson species. Descriptions for nymphs of some other species of chewing lice parasitic on birds are available (e.g., Ansari 1954; Agarwal 1967; Modrzejewska and Złotorzycka 1987; Abrahamovich and Cicchino 1988; Mey 1994; Cicchino and Mey 2007; Agarwal *et al.* 2011; Singh *et al.* 2018, 2019), but to date, their scope and diversity have limited such broad analyses as undertaken by Price and Hellenthal (1996) and Page *et al.* (1995) for mammal hosts.

Mulcticola macrocephalus is the first species of the genus for which all preimaginal stages are fully described. First-instar nymphs have no tergal or sternal plates on the abdomen, second-instar nymphs present roughly rounded lateral and medial tergal plates, whereas third nymphs are similar to second instars but with distinct sternal plates in each side of segments III–VIII. Regarding their growth, we had fewer specimens for statistical analyses than employed by Kéler (1952) and Lonc and Modrzejewska (1986, 1989). However, the analysis of the ratio between the length and width of constant structures such as head and whole body show results similar to those achieved by those authors (Kéler 1952; Lonc and Modrzejewska 1986, 1989). Our plot of these ratios (Fig. 5) confirms that the dimensions of the head and body of chewing lice are good diagnostic characters to determine each nymphal stage (Lonc and Modrzejewska 1986, 1989; Modrzejewska and Złotorzycka 1987). In *M. macrocephalus*, body length increases in greater proportion to corresponding abdominal width during overall growth, whereas the temporal width of the head increases in greater proportion to head length (Fig. 5).

Our results support the ontogenetic classification proposed by Mey (1994) and revised by Smith (2001). The authors classify the nymphal stages in three different groups, based on the morphology of the clypeal region. Species of *Mulcticola* could be classified as part of the group C (*sensu* Mey, 1994), in which all the nymphal instars possess noncircumfasciated heads, with a medially interrupted marginal carina and a dorsal anterior head plate. This characteristic suggests that *Mulcticola* species are part of a more derived group within Philopteridae *s.l.*

In the present paper, we define for the first time the pattern of development for abdominal sclerotisation for a *Mulcticola* species. The only information known on the abdominal structures of the preimagos of this genus confirms Mey's (1994) conclusion. The first nymphal stages lack tergal sclerotisation, and nymphs II and III have two independent plates on each abdominal segment (Valim and Kuabara 2015). The ontogenetic path for these abdominal plates corresponds to the group IIb (*sensu* Mey, 1994).

Descriptions are needed for nymphs from a greater diversity of genera and species from all parts of the world before their morphological characters can become integrated into higher levels of phylogenetic analysis for the Phthiraptera. We intend our description of the nymphs of *M. macrocephalus* to encourage other researchers and contribute to such analyses.

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References

- Abrahamovich, A.H. and Cicchino, A.C. 1988. Estudios bioecologicos, sistematicos y filogeneticos de los malofagos parasitos de *Guira guira* Gmelin (Aves, Cuculidae): III. Diferenciacion de los estadio ninfales de *Vernoniella bergi* (Kellogg, 1906) (Phthiraptera, Philopteridae). *Iheringia Série Zoologia*, **68**: 117–127.
- Agarwal, G.P. 1967. Studies on the bionomics and life-history of *Falcolipeurus frater* (Giebel: 1874) (Mallophaga: Ischnocera). *Indian Journal of Zoology*, **8**: 21–40.
- Agarwal, G.P., Ahmad, A., Rashmi, A., Arya, G., Bansal, N., and Saxena, A.K. 2011. Bio-ecology of the louse, *Upupicola upupae*, infesting the Common Hoopoe, *Upupa epops*. *Journal of Insect Science*, **11**: Article 46, 9 pp. <https://doi.org/10.1673/031.011.4601>.
- Alarie, Y. 1991. Description of larvae of 17 Nearctic species of *Hydroporus* Clairville (Coleoptera: Dytiscidae: Hydroporinae) with an analysis of their phylogenetic relationships. *The Canadian Entomologist*, **123**: 627–704. <https://doi.org/10.4039/Ent122985-9>.
- Ansari, M.A.R. 1954. Pre-imaginal instars of Mallophaga and application of some growth principles. *Pakistan Journal of Science Research*, **6**: 155–161.
- Burmeister, H. 1838. Mallophaga Nitzsch. *Handbuch der Entomologie*, Berlin, **2**: 418–443.
- Carriker, M.A., Jr. 1945. Neotropical Mallophaga miscellany. No. 1. New species of *Struthiolipeurus*, *Mulcticola*, *Microtenia* [sic] and *Pseudocophorus*. *Boletin de Entomologia Venezolana*, **4**: 165–189.
- Carriker, M.A., Jr. 1947. Neotropical Mallophaga miscellany. No. 2. The genus *Ibidoecus* Cummings. *Boletin de Entomologia Venezolana*, **6**: 111–136.
- Carriker, M.A., Jr., 1957. Notes on some of the Vernon L. Kellogg types of Mallophaga. *Microentomology*, **22**: 95–110.

- Cicchino, A.C. and Mey, E. 2007. On morphology, taxonomy, ecology, and distribution of *Bothriometopus* Taschenberg (Phthiraptera, Ischnocera, Philopteridae *sensu lato*). *Rudolstädter naturhistorische Schriften*, **14**: 43–60.
- Clay, T. and Meinertzhagen, R. 1938. New genera and species of Mallophaga. *The Entomologist*, **71**: 275–279.
- Cook, E.F. and Beer, J.R. 1959. The immature stages of the genus *Hoplopleura* (Anoplura: Hoplopleuridae) in North America, with descriptions of two new species. *Journal of Parasitology*, **45**: 405–416, 4 plates.
- Emerson, K.C. 1960. Notes on the Osborn Mallophaga types. *Proceedings of the Biological Society of Washington*, **73**: 155–166.
- Emerson, K.C. 1972. Suborder Ischnocera. *Deseret Test Center, Dugway Proving Ground, Dugway, Utah*.
- Emerson, K.C. and Elbel, R.E. 1957. Two new species of *Mulcticola* (Philopteridae: Mallophaga). *The Canadian Entomologist*, **89**: 420–423. <https://doi.org/10.4039/Ent89420-9>.
- Ferris, G.F. 1951. The sucking lice. *Memoirs of the Pacific Coast Entomological Society*, volume 1. San Francisco, California, United States of America.
- Galloway, T.D. 2006. Ectoparasites (Phthiraptera: Philopteridae; Acari: Ixodidae) of common nighthawk, *Chordeiles minor*, and whip-poor-will, *Caprimulgus vociferus* (Caprimulgiformes: Caprimulgidae), in Manitoba. *Journal of the Entomological Society of Ontario*, **137**: 5–11.
- Galloway, T.D. and Lamb, R.J. 2015. Abundance and stability of populations of a chewing louse, *Mulcticola macrocephalus* (Phthiraptera: Philopteridae), on common nighthawks, *Chordeiles minor* (Caprimulgiformes: Caprimulgidae) in Manitoba, Canada. *The Canadian Entomologist*, **147**: 723–731. <https://doi.org/10.4039/tce.2014.85>.
- Haeckel, E. 1896. *Systematische Phylogenie der wirbellosen Thiere (Invertebrata): Entwurf eines natürlichen Systems der Organismen auf Grund ihrer Stammesgeschichte*. Georg Reimer, Berlin, Germany.
- Hopkins, G.H.E. and Clay, T. 1952. A check list of the genera and species of Mallophaga. *British Museum of Natural History, London, United Kingdom*. 362 pp.
- Kéler, S. von. 1952. Über die Wachstums-Progression bei *Pseudomenopon rowanae* Kéler (Mallophaga). *Beiträge zur Entomologie*, **2**: 113–119. <https://doi.org/10.21248/contrib.entomol.2.1.113-119>.
- Kellogg, V.L. 1896. New Mallophaga, II, from land birds; together with an account of the mallophagous mouth-parts. *Proceedings of the California Academy of Sciences (Series 2)*, **6**: 431–548.
- Kim, K.C. 1965. A review of the *Hoplopleura hersperomydis* complex. *Journal of Parasitology*, **51**: 871–887.
- Kim, K.C. 1966a. The nymphal stages of three North American species of the genus *Enderleinellus* Fahrenholz. *Journal of Medical Entomology*, **2**: 327–330. <https://doi.org/10.1093/jmedent/2.4.327>.
- Kim, K.C. 1966b. The species of *Enderleinellus* (Anoplura, Hoplopleuridae) parasitic on the Sciurini and Tamiasciurini. *Journal of Parasitology*, **52**: 988–1024.
- Lonc, E. and Modrzejewska, M. 1986. An attempt to identification of nymphal instars of biting lice (Mallophaga) using discriminant function. *Polskie Pismo Entomologiczne*, **56**: 153–162.
- Lonc, E. and Modrzejewska, M. 1989. Growth rules applied to the stage identification of nymphal instars of some mallophagan species (Phthiraptera). *Deutsche entomologische Zeitschrift*, **36**: 121–126. <https://doi.org/10.1002/mmnd.19890360118>.
- Mey, E. 1994. Beziehungen zwischen Larvamorphologie und Systematic der Adulti bei den Vogel-Ischnozeren (Insecta, Phthiraptera, Ischnocera). *Mitteilungen aus dem Zoologischen Museum in Berlin*, **70**: 3–84.
- Modrzejewska, M. and Złotorzycka, J. 1987. Studies on the morphology of nymphs of selected Amblycera and Ischnocera Mallophaga. *Polski Pismo Entomologiczne*, **57**: 657–672.

- Nilsson, A. 1989. Larvae of northern European *Hydroporus* (Coleoptera: Dytiscidae). Systematic Entomology, **14**: 99–115. <https://doi.org/10.1111/j.1365-3113.1989.tb00267.x>.
- Osborn, H. 1896. Insects affecting domestic animals. Chapter V. Suborder Mallophaga. In Insects affecting domestic animals: an account of the species of importance in North America, with mention of related forms occurring on other animals. United States Department of Agriculture, Division of Entomology. Pp. 189–249.
- Page, R.D.M., Price, R.D., and Hellenthal, R.A. 1995. Phylogeny of *Geomydoecus* and *Thomomydoecus* pocket gopher lice (Phthiraptera: Trichodectidae) inferred from cladistics analysis of adult and first instar morphology. Systematic Entomology, **20**: 129–143. <https://doi.org/10.1111/j.1365-3113.1995.tb00087.x>.
- Palma, R.L. 1978. Slide mounting of lice: a detailed description of the Canada Balsam technique. New Zealand Entomologist, **6**: 432–436. <https://doi.org/10.1080/00779962.1978.9722313>.
- Pratt, H.D. and Karp, H. 1953. Notes on the rat lice, *Polyplax spinulosa* (Burmeister) and *Hoplopleura oenomydis* Ferris. Journal of Parasitology, **39**: 495–504.
- Price, R.D. and Hellenthal, R.A. 1996. Taxonomic importance of first-instar chewing lice (Phthiraptera: Trichodectidae) from pocket gophers (Rodentia: Geomyidae). Annals of the Entomological Society of America, **89**: 510–518. <https://doi.org/10.1093/aesa/89.4.510>.
- Price, R.D., Hellenthal, R.A., and Palma, R.L. 2003. World checklist of chewing lice with host associations and keys to families and genera. In The chewing lice: world checklist and biological overview. Edited by R.D. Price, R.A. Hellenthal, R.L. Palma, K.P. Johnson, and D.H. Clayton. Illinois Natural History Survey Special Publication, **24**: 1–448.
- Singh, P., Gupta, N., Khan, G., Kumar, S., and Ahmad, A. 2019. Diagnostic characters of three nymphal instars and morphological features of adult Collard-dove louse *Columbicola bacillus* (Phthiraptera: Insecta). Journal of Applied and Natural Science, **11**: 7–11. <https://doi.org/10.31018/jans.v11i1.1855>.
- Singh, P., Khan, G., Kumar, S., and Gupta, N. 2018. Morphological features of adults and nymphal instars of *Coloceros* (sic) species (Phthiraptera: Insecta) recovered from Collared-dove *Streptopelia decaocto*. Journal of Applied and Natural Science, **10**: 1341–1344. <https://doi.org/10.31018/jans.v10i4.1853>.
- Smith, V.S. 2001. Avian louse phylogeny (Phthiraptera: Ischnocera): a cladistic study based on morphology. Zoological Journal of the Linnean Society, **132**: 81–144. DOI: [10.1046/j.1365-3113.2000.00095.x](https://doi.org/10.1046/j.1365-3113.2000.00095.x).
- Valim, M.P. and Kuabara, K.M.D. 2015. The feather louse genus *Mulcticola* Clay et Meinertzhagen, 1938 (Phthiraptera: Philopteridae) from Brazil, with descriptions of five new species and catalogue for species described in the genus. Folia Parasitologica, **62**: 1–15. <https://doi.org/10.14411/fp.2015.036>.